

Brief Note

Effects of Prostaglandin A₂ on the Short-Circuit Current across the Isolated Rabbit Iris-Ciliary Body

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The ionic composition of the aqueous humor is characterized by a higher concentration of HCO₃⁻ and lower concentrations of Na⁺ and K⁺ than either serum or cerebrospinal fluid. To explain the specific ion composition, active ion transport mechanisms in the ciliary epithelial cells have been postulated by several authors.¹⁻⁴⁾ Since water moves accompanied by electrolytes, the disturbance of ion transport mechanisms may possibly produce abnormal intraocular pressure such as that observed in glaucoma. Prostaglandins (PGs) have been known to affect ion transport^{3,4)} and they have been practically used to lower intraocular pressure.

In my previous work,⁵⁾ it was demonstrated that PGF_{2α} and PGD₂ increased the short-circuit current (SCC) in the isolated rabbit iris-ciliary body by modification of anion transport rather than Na⁺-K⁺ active transport. The present work was carried out to study the action of PGA₂ in comparison with those of PGF_{2α} and PGD₂, because PGA₂ is known to be most effective for the treatment of intraocular pressure elevation.⁶⁾

The method used for the measurement of SCC has been described in my previous paper,⁵⁾ but a slight improvement was made in the method for mounting the iris-ciliary body preparation on the Ussing-type chamber. A nylon sheet was stretched tightly over the silicone platform with a small hole of 2.1 mm diameter or 0.035 cm² area. The preparation was placed in such a way that the ciliary body would be centered over the small hole. A second nylon sheet was gently placed over the preparation, as illustrated in Fig. 1. An adhesive liquid was used to avoid slipping of the preparation from the silicone platform. The layer of the silicone, nylon sheets and preparation was placed between two plastic plates. The completed preparation-mounting block was then placed between the two halves of the Ussing-type chamber. This improvement in mounting of the preparation helped to minimize the leakage of current. The standard solution contained 118 NaCl, 4.7 KCl, 0.5 KH₂PO₄, 1.2 MgCl₂, 2.5 CaCl₂, 15 HEPES, 5.5 glucose and 15.5 NaHCO₃ (mM) with pH = 7.4. The solution was maintained at 36°C and was bubbled continuously with a mixture of 95% O₂ and 5% CO₂.

The experiment was commenced about one hour after mounting of the preparation. Upon closing the feedback circuit, a current of 0.3-0.7 μA flowing from the stromal side to the aqueous side was recorded. This is the background

or basal SCC. Since the area of the fenestration was 0.035 cm^2 , it corresponded to $8.5\text{--}20 \mu\text{A}/\text{cm}^2$. The smaller value of the background SCC than that recorded in the previous work was the result of the better mounting.

When $50 \mu\text{l}$ of 10^{-3} M PGA_2 solution was added to the 5 ml chamber facing the aqueous side, SCC started to increase gradually and attained a steady value in more than 60 min (Fig. 2). Contrary to the findings on $\text{PGF}_{2\alpha}$ or

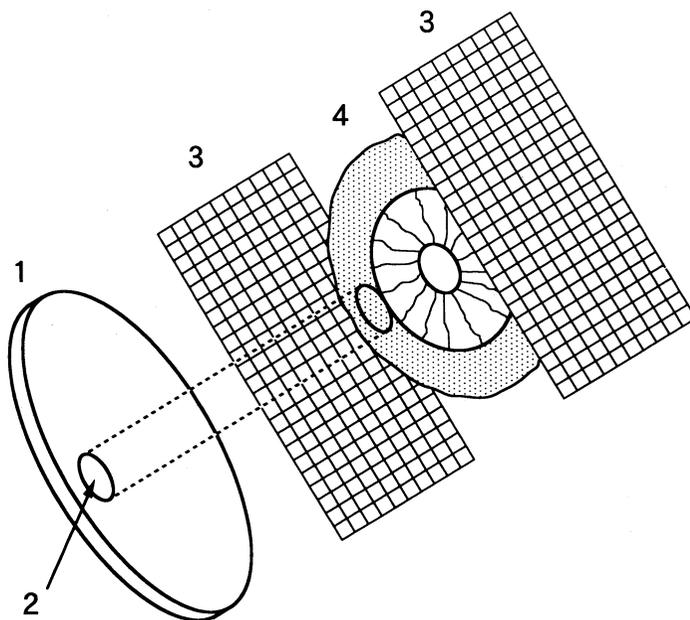


Fig. 1. Schematic diagram of the mounting method for the iris-ciliary body preparation. 1: Silicone plate, 2: Fenestration of an area of 0.035 cm^2 , 3 and 4: Nylon nets adhering to the iris-ciliary body.

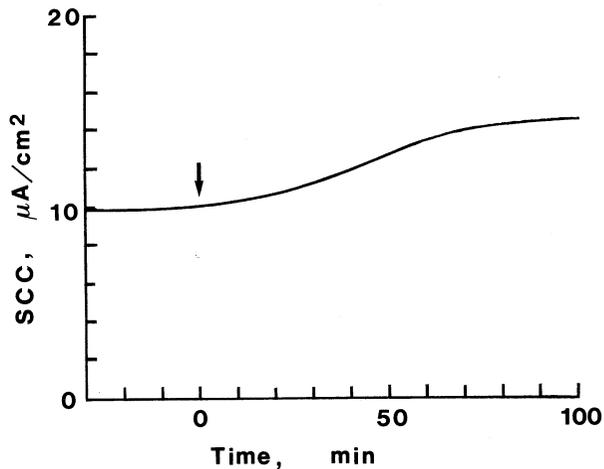


Fig. 2. Effects of PGA_2 ($1 \times 10^{-5} \text{ M}$) on the SCC. At the time indicated by the arrow, 0.05 ml of 10^{-3} M PGA_2 was applied to the 5 ml chamber facing the humoral side.

PGD₂ in the previous work, the time course of the increase of SCC caused by PGA₂ was very slow. Delay due to diffusion for added PGA₂ to reach the preparation was excluded because the fluid in the chamber was continuously stirred by bubbling with a gas. Addition of PGA₂ to the chamber of the stromal side resulted in no significant changes in SCC. In Table 1, the increases in SCC caused by PGA₂ obtained from four preparations are summarized together with the results for PGF_{2α} and PGD₂ obtained in my previous work. The increase in SCC caused by 10⁻⁵ M PGA₂ was smaller and weaker than that in PGF_{2α} at the same concentration. Namely, PGA₂ was less potent than PGF_{2α}. On the other hand, the effective concentration of PGA₂ to produce a considerable increase in SCC was 1×10⁻⁵ M, whereas that of PGD₂ was 1×10⁻⁴ M. In this regard, PGA₂ was more effective than PGD₂. In summary, the order of the action was PGF_{2α} > PGA₂ > PGD₂.

TABLE 1. Changes in the amplitude of SCC ($\mu\text{A}/\text{cm}^2$) caused by PGF_{2α}, PGD₂ and PGA₂ added to the aqueous side

	control	test	% ratio of increase
PGF _{2α}	17.0	19.5	27.6±12.8 (SE)
	19.5	26.3	
10 ⁻⁵ M	20.3	27.0	
PGD ₂	12.8	14.7	38.8±29.3
	11.9	20.0	
	12.6	18.6	
10 ⁻⁴ M	12.3	15.1	
PGA ₂	9.6	10.9	23.8±19.1
	9.8	14.0	
10 ⁻⁵ M	15.9	18.0	
	16.4	20.8	

The nature of the increase in SCC can be explained by inhibition of the Na⁺-K⁺ pump of the epithelial cell located on the aqueous side or by facilitation of Na⁺-HCO₃⁻ cotransport existing on the same side of the cell. Na⁺-H⁺ or HCO₃⁻-Cl⁻ antitransport may be involved in the change in SCC. Facilitation of either the Na⁺-K⁺ pump or Na⁺-HCO₃⁻ cotransport would result in an increase in water secretion and a rise in the intraocular pressure. This is contrary to the explanation for application of PGs to practical use. Probably, PGs lower the intraocular pressure by dilating the canal of Schlemm to promote outflow of aqueous humor *in situ*.⁷⁾ If PGs increase the production of aqueous humor from the epithelial cells and also promote outflow into the canal of Schlemm, their action on the intraocular pressure stands on the balance of two opposite effects. It has been reported that PGE₂ and PGF_{2α} caused an initial rise preceding a sustained fall of the intraocular pressure when applied through an eye dropper.⁸⁾ The initial rise may be due to the strong action on aqueous humor production. On the contrary, PGA₂ and PGD₂ are known to bring about a long-lasting fall in intraocular pressure without an initial rise. The mild effect of PGA₂ on the ion transport and the subsequent water secretion may be advantageous for its practical use to lower the intraocular pressure.

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